



Build NEURON microcircuits using touch detection with Snudda

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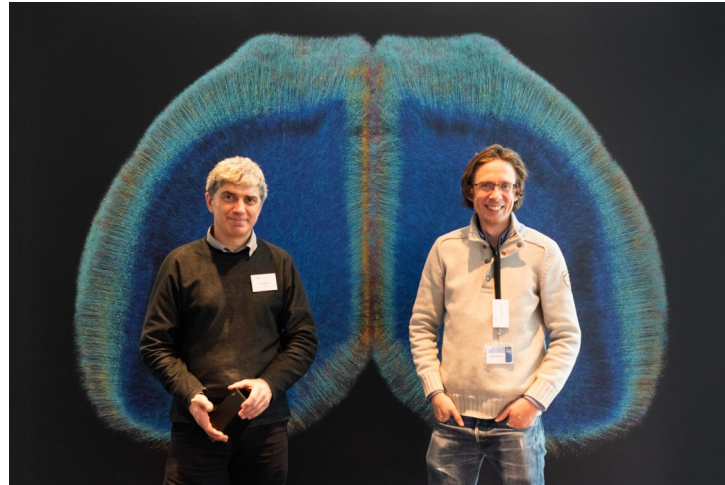
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Swedish e-Science Research Centre

dBrain

Hjorth, Kozlov et al, 2020. The microcircuits of striatum in silico. PNAS <https://doi.org/10.1073/pnas.2000671117>

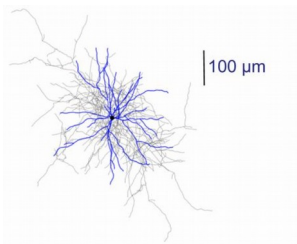
Hjorth, Hellgren Koteleski, Kozlov 2021. Predicting Synaptic Connectivity for Large-Scale Microcircuit Simulations Using Snudda Neuroinformatics <https://doi.org/10.1007/s12021-021-09531-w>

Frost Nylén, Hjorth, Grillner, Hellgren Koteleski 2021. Dopaminergic and Cholinergic Modulation of Large Scale Networks in silico Using Snudda <https://dx.doi.org/10.3389%2Fncir.2021.748989>

Model building pipeline

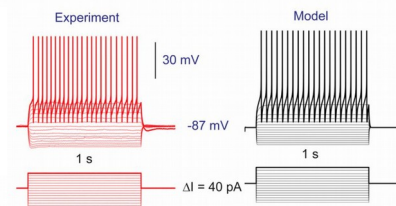
Morphology repair

Treem



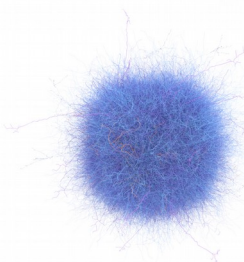
Model optimisation

BluePyOpt



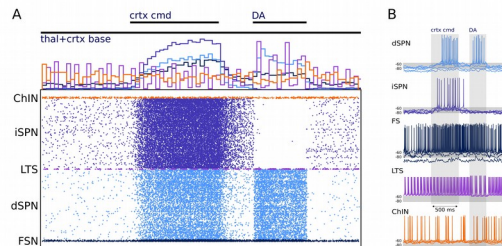
Network creation

Snudda



Simulation

NEURON



Open Source

<https://github.com/a1eko/treem>
<https://github.com/BlueBrain/BluePyOpt>
<https://github.com/Hjorthmedh/Snudda>
<https://github.com/neuronsimulator/nrn>

What is Snudda?

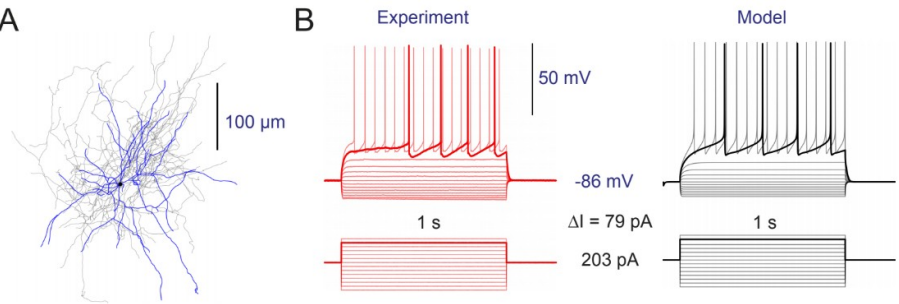
- Voxel-based touch detection
 - Rule-based pruning of synapses
 - Network analysis
 - Simulation of network
-
- Open source -- <https://github.com/hjorthmedh/Snudda>
 - Runs on Linux, Mac and Windows desktops and on super computers



Components needed

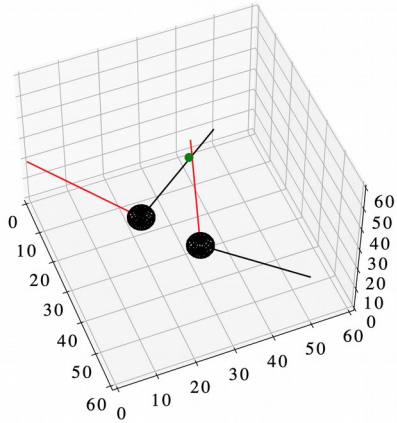
Requires:

- Complete morphologies with dendrites, ^A and preferably also axons
- Detailed electrophysiological neuron models optimised using BluePyOpt
- Synapse models
- Connectivity statistics of the network

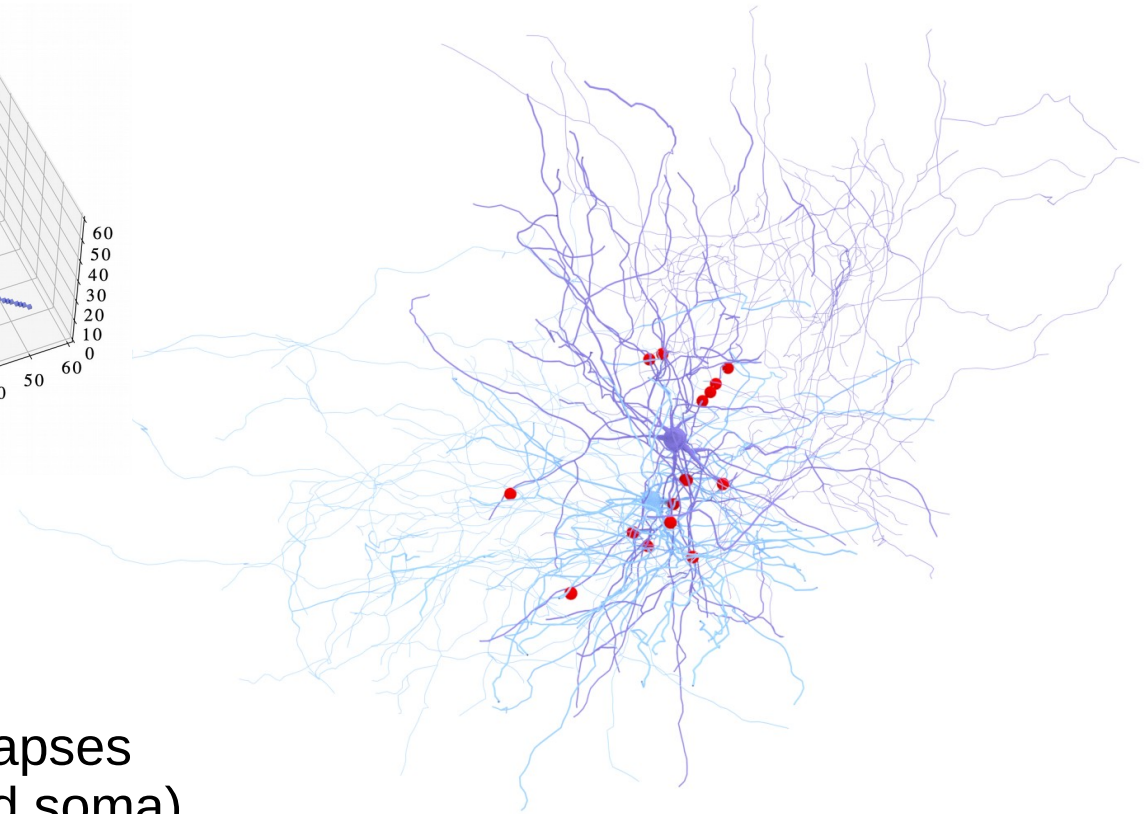
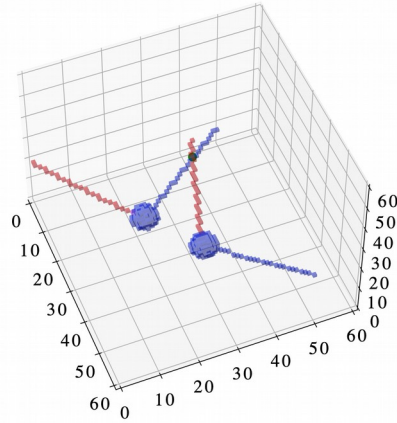


Voxel based touch detection

A

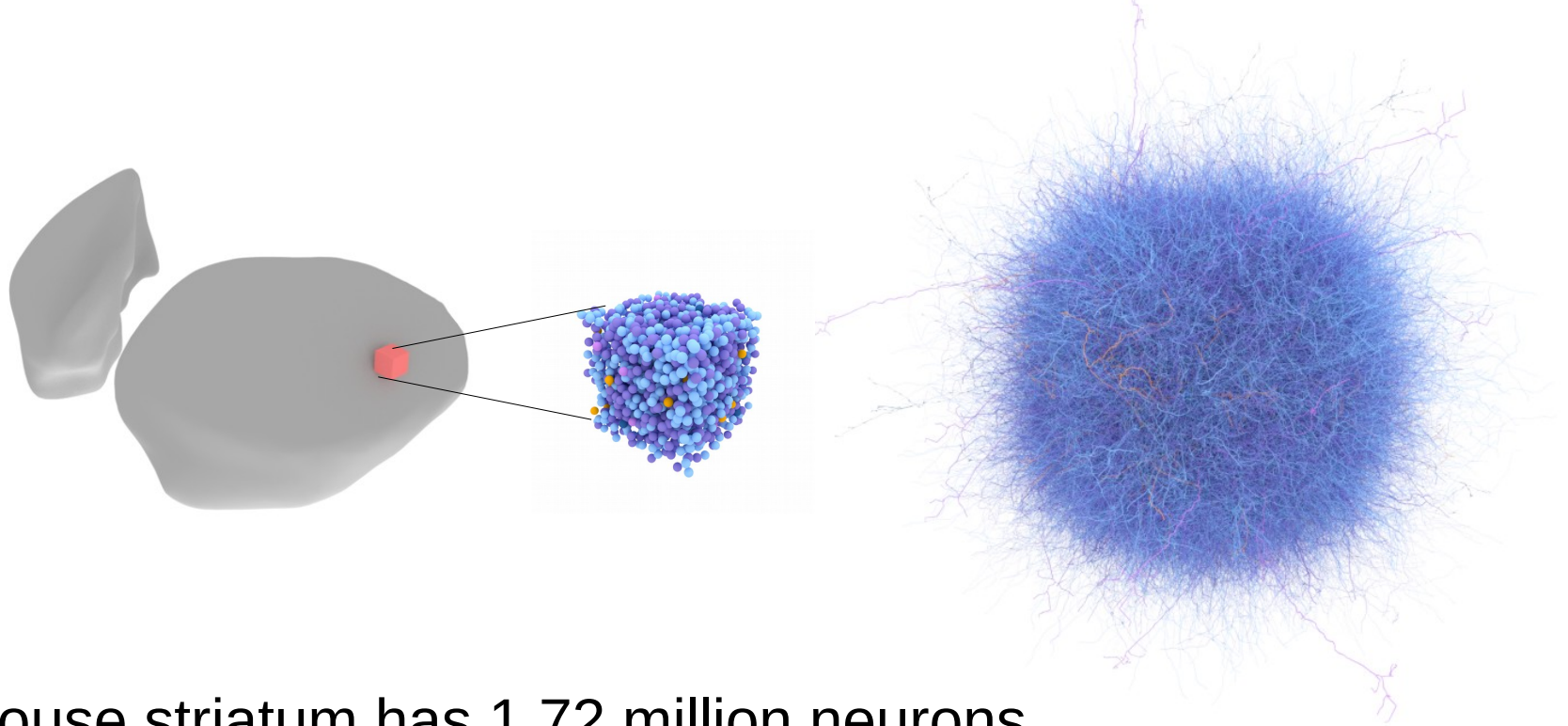


B



Space divided into voxels, $3\mu\text{m}$ side

Touch detection places putative synapses
where axons are near dendrites (and soma)



Entire mouse striatum has 1.72 million neurons

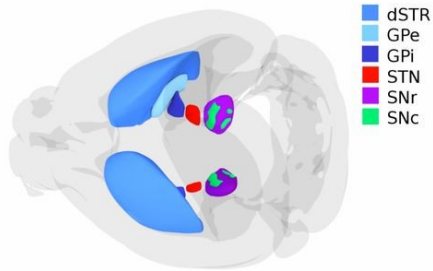
We detect 31 billion putative synapse locations

All locations does not have synapses -- we need to prune!

Basal Ganglia

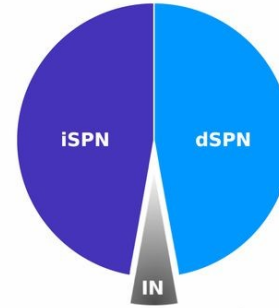
A

Basal ganglia



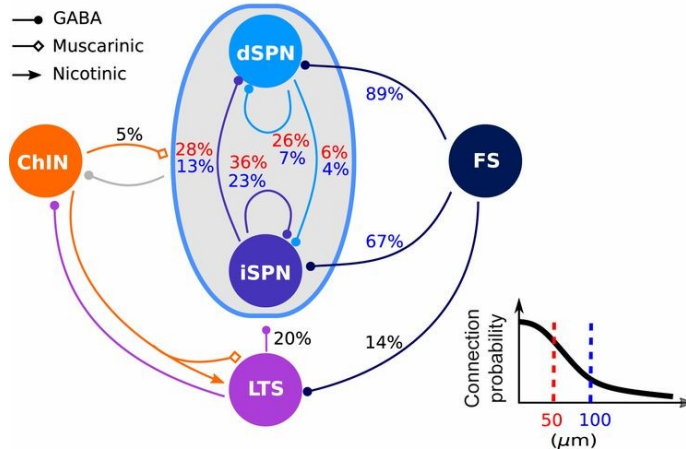
B1

Cell type distribution in the striatum

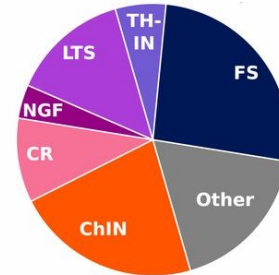


C

Striatal microcircuit

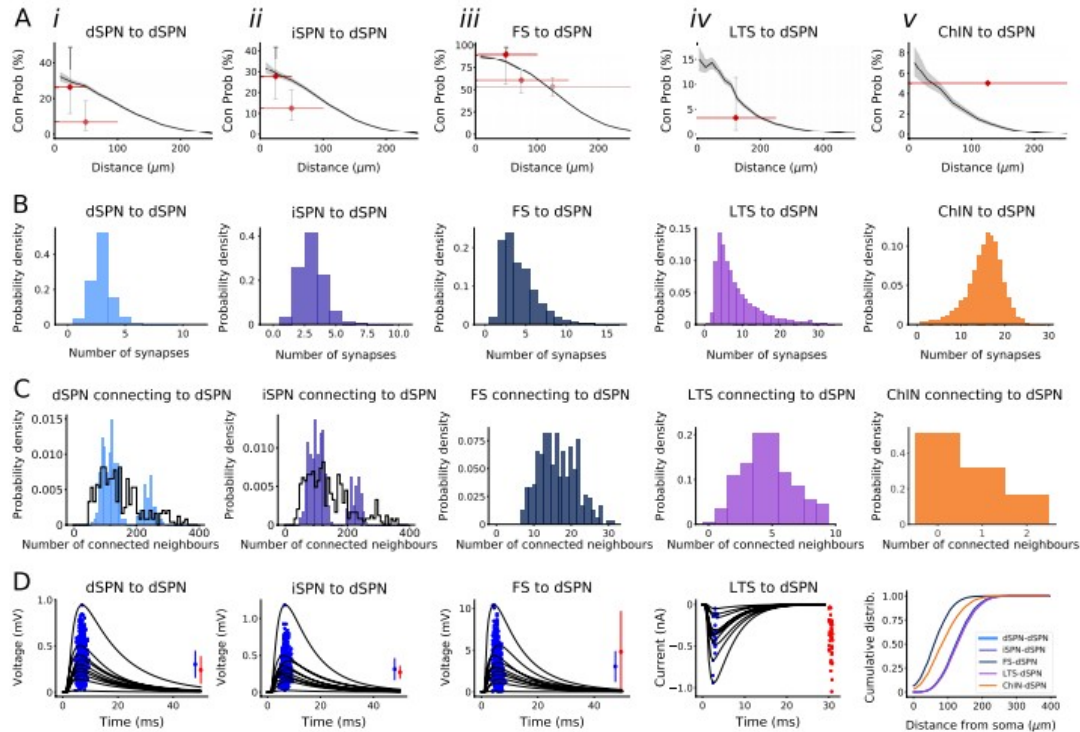


B2

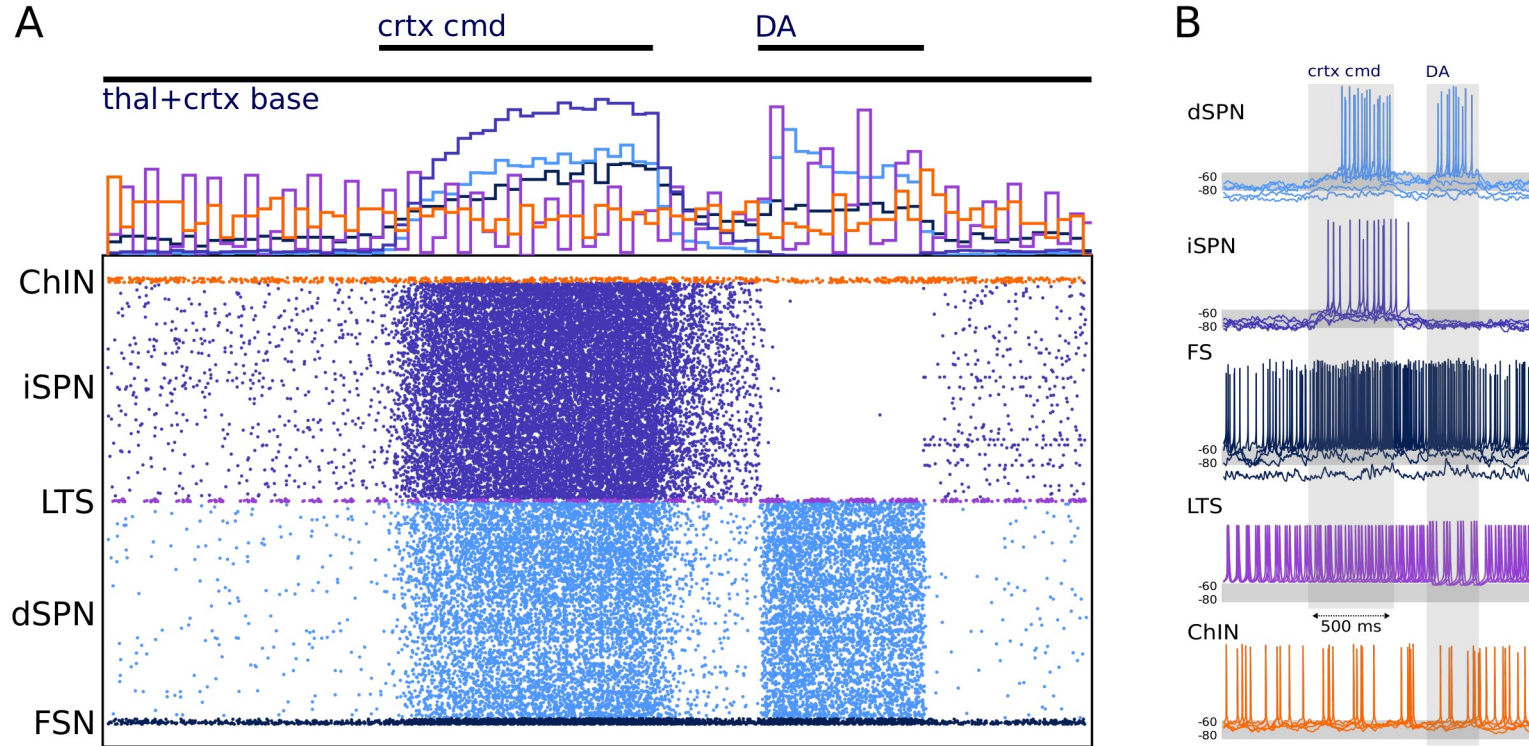


Interneurons

Prune synapses to match experimental pair-wise connection probability



Simulation of Striatum



Snudda

> pip install snudda

Repository:

<https://github.com/Hjorthmedh/Snudda>

Jupyter Notebook:

> pip install notebook

> jupyter notebook



<https://github.com/Hjorthmedh/Snudda/tree/master/examples/notebooks>

Jupyter Notebooks on Github

<https://github.com/Hjorthmedh/Snudda/tree/master/examples/notebooks>

Simple network creation

This example creates a striatal network of 100 dSPN and 100 iSPN neurons. You can also do this using the command line interface, see "snudda -h".

This notebook is started in the `Snudda/snudda/examples/notebooks` directory, as all paths are given relative to there.

First we create a `network-config.json` file in `networks/simple_example` which is a subdirectory to the `notebooks` directory. The 200 neurons are placed inside a cube, with cell density 80500 neurons/mm³. The neuron morphologies and parameters are taken from the `Snudda/snudda/data/neurons/dspn` and `ispn` folders.

Here we have set the `random_seed` to 12345.

```
In [1]: import os
        from snudda import SnuddaInit

network_path = os.path.join("networks", "simple_example")
config_file = os.path.join(network_path, "network-config.json")
cnc = SnuddaInit(config_file=config_file, random_seed=12345)
cnc.define_striatum(num_dSPN=100, num_iSPN=100, num_FS=0, num_LTS=0, num_ChIN=0,
                  volume_type="cube", neurons_dir="$DATA/neurons")
cnc.write_json(config_file)
```

Using cube for striatum

Adding neurons: dSPN from dir \$DATA/neurons/striatum/dspn

Adding neurons: iSPN from dir \$DATA/neurons/striatum/ispn

Writing networks/simple_example/network-config.json

This reads in the `network-config.json` file and places the dSPN and iSPN neurons within the cube volume, then writes the positions to the `network-neuron-positions.hdf5` file.

```
In [2]: from snudda import SnuddaPlace
        sp = SnuddaPlace(network_path=network_path)
```

Notebook examples for Snudda

Here is a collection of Jupyter Notebooks, some of the workflows are split over multiple notebooks, and you have to run them in a specific order. Snudda uses HDF5 files and they can sometimes be locked by one Notebook, so make sure to shutdown the kernel in the old Notebook before proceeding to the next one.

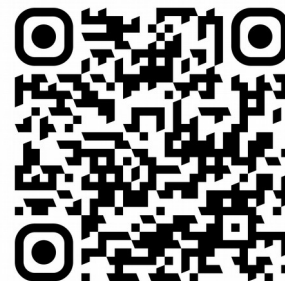
Network creation

- [simple_network_creation](#) first example of network creation
- [analyse_network_connectivity](#) analyses network connectivity created by [simple_network_creation](#).
- [simple_network_parallel](#) how to use ipyparallel when running Snudda
- [custom_slice_example](#) shows how to create custom slice and define your own connectivity rules for neuron types.
- [population_unit_network](#) how to define population units.
- [example_of_density_function](#) how to specify density variations using a function of (x,y,z) in a volume.
- [example_of_neuron_rotations](#) shows how to rotate neurons based on position.
- [connect_structures_example](#) shows how to create neuron projections between volumes when no-axon data is available ([parallel version](#)).

Input creation

- [input_generation_example_1](#) generate constant Poisson input (uses [simple_network_creation](#))
- [input_generation_example_2_frequency_vectors](#) define Poisson input with multiple start/stop times (uses [simple_network_creation](#)).
- [input_generation_example_3_correlation](#) finer control in input targeting (uses [population_unit_network](#))
- [input_tuning_example](#) explore what input number and frequency are good neurons, e.g to avoid depolarisation block.

Striatum example



<https://github.com/Hjorthmedh/Snudda/tree/master/examples/notebooks>

Input creation

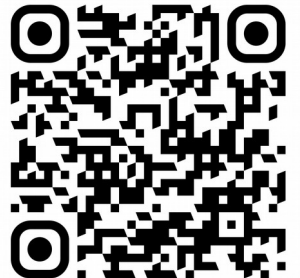
- [input_generation_example_1](#) generate constant Poisson input (uses [simple_network_creation](#))
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- [input_tuning_example](#) explore what input number and frequency are good neurons, e.g to avoid depolarisation block.

🔗 Striatum example

- [striatum_example](#) creates a small striatal network, increase number of neurons for the full version.
- [striatum_example_simulate](#) sets up input and simulates the [striatum_example](#) network.
- [striatum_example_plot](#) plots spike raster from [striatum_example_simulate](#). Figure 4 in Methods paper has a larger version.

Visualisation

- [blender_example](#) - Use Blender to visualise a network, or part of a network
- [Hjorth, Hellgren Kotaleski, Kozlov](#), Neuroinformatics 2021 figures



To run in parallel

```
> export IPYTHONDIR="`pwd`/.ipython"
> export IPYTHON_PROFILE=Snudda_LOCAL

> ipcluster start -n 4 --profile=$IPYTHON_PROFILE --ip=127.0.0.1&
> sleep 20
> simName=networks/mynetwork
> snudda init $simName --size 10062 --overwrite
> snudda place $simName --parallel
> snudda detect $simName --parallel
> snudda prune $simName --parallel

> cp -a data/input-config/input-tinytest-v9-freq-vectors.json $simName/input.json
> snudda input $simName --parallel
> ipcluster stop
> mpiexec -n 6 snudda simulate $simName
```

